

# Manganese(III) Complexes of Bis(hydroxyphenyl)dipyrromethenes Are Potent Orally Active Peroxynitrite Scavengers

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#### S Supporting Information

ABSTRACT: We report a new series of biscyclohexanofused Mn(III) complexes of bis(hydroxyphenyl)dipyrromethenes, 4a-c, as potent and orally active peroxynitrite scavengers. Complexes 4a-c are shown to reduce peroxynitrite through a two-electron mechanism, thereby forming the corresponding Mn(V)O species, which were characterized by UV, NMR, and LC-MS methods. Mn(III) complex 4b and its strained BODIPY analogue 9b were analyzed by X-ray crystallography. Finally, complex 4a is shown to be an orally active and potent analgesic in a model carrageenaninduced hyperalgesia known to be driven by the overproduction of peroxynitrite.

The overproduction of reactive oxygen species (ROS) in vivo **I** is now widely recognized as a key contributor to numerous pathologies.<sup>1</sup> One particularly damaging situation results from the diffusion-controlled radical coupling of the central ROS, superoxide, with nitric oxide to form peroxynitrite.<sup>2</sup> The highly reactive peroxynitrite is a powerful biological oxidant that leaves a trail of dysfunctional oxidized and nitrated proteins, lipids, and nucleotides in its wake.<sup>3</sup> From a pharmacological perspective, peroxynitrite is considered a potent proinflammatory and proapoptotic species that plays a critical role in pain of several etiologies, as demonstrated initially by our team and then by others.<sup>4-6</sup> Accordingly, the discovery of pharmaceutically relevant agents that can effectively decompose peroxynitrite should have significant therapeutic value.<sup>2,3</sup>

As a result of the early discoveries of Groves<sup>7</sup> and Stern,<sup>8</sup> Mn(III) and Fe(III) porphyrins have emerged as important classes of peroxynitrite reductase and isomerase catalysts, respectively (Figure 1A). Elegant mechanistic studies have revealed that the more pharmacologically suitable Mn(III) porphyrins decompose peroxynitrite primarily in a one-electron fashion and require a biological coreductant such as ascorbate to complete the reductase catalytic cycle.<sup>9</sup> One-electron reduction of peroxynitrite produces the potentially damaging nitrogen dioxide radical, which is also thought to undergo rapid reduction by endogenous antioxidants.<sup>5</sup> Thus, if endogenous antioxidants are plentiful, Mn(III) pophryins can fully detoxify peroxynitrite in vivo. From this class, the isomeric Mn(III) tetrakis(meso-Nalkylpyridinium)porphyrins [e.g.,  $Mn(III)-4-TMPyP^{5+}$  (1)]



Figure 1. (A) Peroxynitrite decomposition catalysts 1 and 2. (B) Bis(hydroxyphenyl)dipyrromethene analogues 3 and 4.

are the most well studied, both as peroxynitrite reductase catalysts and superoxide dismutase (SOD) mimics.<sup>9,10</sup>

Recently, Gross has reported that Mn(III) and Fe(III) corroles are also excellent peroxynitrite decomposition catalysts.<sup>11</sup> Remarkably, the Mn(III) corroles operate through a two-electron cycle, reducing peroxynitrite to nitrite instead of nitrogen dioxide through a novel disproportionation mechanism. The most important finding from that work was that Mn(III) corroles can decompose peroxynitrite in a catalytic fashion [in contrast to Mn(III) porphyrins] and therefore do not require the assistance of endogenous coreductants.

Although Mn(III) porphyrins such as 1 and Mn(III) corrole systems such as 2 have proven to be powerful pharmacological tools in animal studies, where they have demonstrated the benefits of destroying peroxynitrite in vivo,  $^{11-14}$  they are not optimal as therapeutic candidates. While these types of polycationic complexes have excellent catalytic activities and their high water solubility is useful for laboratory measurements, their corresponding high polarity renders them poorly membrane-soluble.

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More amphiphilic metallocorroles have indeed shown great promise as orally active peroxynitrite decomposition catalysts.<sup>12c</sup> Unfortunately, synthetic methods for accessing polyfunctional corrole systems remain quite challenging, and thus, these systems are not particularly suited for iterative structure—activity relationship (SAR) studies. In view of this, we have been keenly interested in the design, synthesis, and evaluation of new catalyst systems with enhanced druglike properties.

In our search for alternative trianionic ligand systems capable of supporting a Mn(V)O intermediate arising from the twoelectron reduction of peroxynitrite [similar to Mn(III) corroles], we were intrigued by the B,O-chelated boron dipyrromethene (BODIPY) dye 3 first reported by Burgess (Figure 1B).<sup>15</sup> This and related dyes are constrained because of the chelate effect and thus have improved fluorescence properties relative to their nonchelated and well-known BODIPY congeners. Not only is the overall "ligand set" in these systems perfect for mimicking the trianionic corrole, but these compounds are also amenable to modular synthesis in good to excellent yields.<sup>16</sup> Here we report the synthesis and evaluation of new biscyclohexano-fused Mn(III) complexes of bis(hydroxyphenyl)dipyrromethenes as potent peroxynitrite scavengers with druglike properties.

The syntheses commenced with readily available tetrahydroisoindole 5, <sup>17</sup> which was converted to Boc-protected 3-bromotetrahydroisoindole 6 in 95% yield over two steps (Scheme 1). Next, compound 6 underwent smooth Suzuki couplings with a representative set of protected hydroxyphenylboronic acids (7a-c) in 70-89% yield, and this was followed by a one-pot decarboxylation/deprotection procedure that furnished the corresponding 2-benzyloxy- or 2-methoxyphenyltetrahydroisoindole derivatives 8a-c. Compounds 8a-c were then treated with benzaldehyde and trifluoroacetic acid (TFA) under Lindsey conditions<sup>18</sup> to form the corresponding dipyrromethane derivatives; this was followed by oxidation to the dipyrromethenes (DIPYs) with *p*-chloranil<sup>19</sup> and subsequent phenol ether deprotec-tion with boron tribromide.<sup>19,20</sup> This sequence was carried out without intervening purifications. The boron tribromide deprotection step afforded the crude BODIPY systems 9a-c, which were then directly converted to the Mn(III) complexes 4a-c by reaction with manganese(II) chloride under basic aerobic conditions. The darkemerald-green complexes 4a-c behave as simple lipophilic "organic" molecules and were thus amenable to purification by flash chromatography on silica gel, which provided analytical materials for characterization and activity studies. The overall yield of this sequence ranged from 31 to 56% (unoptimized) and is therefore quite competitive with porphyrin and corrole syntheses, which require the preparation of functionalized pyrrole and/or dipyrromethane units followed by an often low-yield cyclization/oxidation step.<sup>18</sup>

Magnetic susceptibility measurements confirmed representative complex **4b** to be a high-spin d<sup>4</sup> Mn(III) complex. <sup>1</sup>H NMR spectra of **4a** in CD<sub>2</sub>Cl<sub>2</sub> revealed broad, shifted peaks indicative of the paramagnetic Mn(III) complex. Oxidation of **4a** with *m*-CPBA in CD<sub>2</sub>Cl<sub>2</sub> afforded normal sharpened peaks characteristic of the corresponding diamagnetic low-spin d<sup>2</sup> Mn(V)O complex (Figures S21 and S22 in the Supporting Information).<sup>21</sup>

Treatment of green-colored methanolic solutions of the Mn-(III) complexes 4a-c with excess peroxynitrite (in 0.1N NaOH)<sup>22</sup> afforded the corresponding red-colored Mn(V)O intermediates (Figure 2). In methanol solution, the Mn(V)O species persisted for 20–30 min and were stable enough to be confirmed by LC–MS (Figure 2 inset) and UV–vis spectroscopy. The UV–vis spectral changes are analogous to those observed during the oxidative generation and subsequent reduction of Mn(V)O corroles<sup>23</sup> and Scheme 1. Synthesis of Mn(III) Complexes of Bishydroxyphenyl-DIPYs<sup>a</sup>



<sup>a</sup> Conditions: (a) NBS, THF, 100%; (b)  $Boc_2O$ , 4-DMAP,  $CH_3CN$ , 95%; (c) 5%  $Pd(PPh_3)_4$ ,  $Na_2CO_3/H_2O$ ,  $CH_3OH$ , toluene, 70–89%; (d)  $(CH_2OH)_2$ , KOH, 195 °C, 70–85%; (e) PhCHO, cat. TFA, 97–99%; (f) *p*-chloranil,  $CH_2Cl_2$ ; (g) BBr<sub>3</sub>,  $CH_2Cl_2$ ; (h)  $MnCl_2$ ,  $CHCl_3$ ,  $CH_3OH$ , 2,6-lutidine, air, 68–79% for three steps (from f).

in Goldberg's elegant studies of highly functionalized Mn(V)O corrolazines.<sup>21</sup> For all three complexes  $4\mathbf{a}-\mathbf{c}$ , the oxo species were clearly observable as  $[M + H]^+$  species using positive-ion-mode LC-MS, confirming the formal oxidation state of  $[Mn(V)O]^{3+}$  (accounting for the trianionic ligand set). Treatment of the methanolic solutions of Mn(V)O species with 5 equiv of ascorbate in phosphate buffer (pH 7.2) resulted in the apparent instantaneous conversion back to the green Mn(III) form, effecting a possible reductase mode of action.

The results shown in Figure 2 are of interest only for identifying the putative two-electron oxidation of 4b with subsequent study of the Mn(V)O species **10b** by spectroscopic methods. However, to confirm that the complexes can indeed decompose peroxynitrite more rapidly than its spontaneous decomposition at physiological pH, a relevant rapid-throughput in vitro assay was sought.

Complexes 4a-c were therefore assayed for their peroxynitrite decomposition activity by determining their ability to inhibit aryl boronate oxidation.<sup>24</sup> Oxidation of 4-acetylphenylboronic acid to 4-acetylphenol by peroxynitrite is a clean conversion devoid of any observable intermediates, and the second-order rate constant for this reaction has been accurately measured to be  $k = 1.6 \times 10^{6} \text{ M}^{-1}$ s<sup>-1</sup> using stopped-flow spectrophotometric methods.<sup>24</sup> Inhibition results for complexes 4a-c and 1 are presented in Table 1. From the percent inhibition values, we calculated the corresponding apparent second-order rate constants for oxidation of the Mn(III) form to the Mn(V)O form for 4a-c and the Mn(IV)O form for 1 at 25 °C in phosphate buffer (pH 7.2). The numbers match up well with the rate constants reported in the literature for the well-known complex 1.<sup>25</sup> Thus, this assay provides a reliable method for the rapid in vitro measurement of activity toward decomposition of peroxynitrite prior to in vivo studies. Our data reveal that analogues 4a-c have estimated apparent rate constants in the range of those observed for manganese corrole systems.<sup>11</sup> If the resulting Mn(V)O forms are more rapidly reduced back to the Mn(III) resting state by endogenous reductants, the k values in Table 1 would be the catalytic rate



Figure 2. UV-vis spectral changes for 4b (33  $\mu$ M, MeOH, 25 °C) after treatment with 0–20 equiv of peroxynitrite (spectra were collected every 2 min; see the Supporting Information). Center inset: samples of 4b and 10b. Right inset: LC-MS data ( $[M + H]^+$ ) for 10b.

constants for reductase-type activity.<sup>9</sup> In that manganese corroles have been shown to possess peroxynitrite decomposition activity without the need for endogenous reductants,<sup>11</sup> complexes 4a-c may also operate via a similar catalytic cycle, as they possess a similar trianionic ligand environment. Future mechanistic studies will address this possible mode of catalysis.

Single-crystal X-ray analysis of 4b and its BODIPY analogue, the synthetic precursor 9b, provided very interesting structural information for this new ligand class (Figure 3). The BODIPY system 9b shows significant distortion of the tetrahedral boron center, similar to that observed for the related non-cyclohexano system.<sup>15</sup> In the case of 9b, the O-B-O angle is 108.4°, but the N1-B-O2 and N2-B-O1 angles are 114.4 and 113.1°, respectively. The N–B–N angle is pinched inward to 106.5°, most likely in response to the phenyl group that bisects the planar dipyrromethene unit and therefore feels a close steric interaction with the CH<sub>2</sub> groups of the neighboring fused cyclohexano rings. The Mn(III) complex 4b crystallized with coordinated axial methanol molecules (from the CH2Cl2/MeOH solution) to afford a Jahn-Teller-distorted octahedral array around the manganese atom. The phenyldipyrromethene unit in 4b is displaced upward relative to the equatorial plane, with the coordinating phenolate groups displaced slightly downward. The release in strain associated with the distorted tetrahedron of 9b most likely drives its direct conversion to 4b without the need for isolation of the free phenol ligand. The two axial Mn-O(MeOH) bonds are considerably longer than those in similar structures observed for Mn(III) corroles (2.19 Å)<sup>11c</sup> and Mn(III) corrolazines (2.107 Å),<sup>21b</sup> most likely because of the Jahn-Teller effect.<sup>26</sup> The Mn-O3 bond distance is 2.226 Å and the longer Mn–O4 bond distance is 2.342 Å, possibly also influenced by steric interactions with the saddled hydroxyphenyl groups.

In the well-known Hargreaves model,<sup>28</sup> intraplantar injection of carrageenan leads to the time-dependent development of thermal hyperalgesia in rats, an inflammatory response known to be driven by high levels of peroxynitrite flux (Figure 4).<sup>29</sup> This hyperalgesic effect was potently inhibited when **4a** was given by oral gavage





<sup>*a*</sup> Octanol—water partition coefficients measured using the slow-stir method.<sup>27 *b*</sup> Apparent second-order rate constant for the oxidation of complex (1 equiv) by peroxynitrite (1 equiv) estimated from the % inhibition in 100 mM phosphate buffer (pH 7.2) with no secondary antioxidants added. Inhibition data were determined by LC—MS after 1 min of reaction time. <sup>*c*</sup> Second-order rate constant measured using stopped-flow methods.<sup>25</sup>



Figure 3. Single-crystal structural analysis of BODIPY analogue 9b and Mn(III) complex 4b.

(Figure 4). Nearly 100% inhibition for 2 h was seen, with a substantial inhibitory effect maintained out to 5 h. At the 2 h time point, a similar degree of inhibition was observed with the nonselective COX-1/COX-2 inhibitor ibuprofen at 300 mg/kg ( $99 \pm 5\%$  inhibition, n = 6). Under the same conditions and at 300 mg/kg, acetaminophen or aspirin attenuated hyperalgesia by  $20 \pm 4\%$  and  $50 \pm 6\%$  respectively (n = 6, P < 0.05 relative to carrageenan alone at the 2 h time point). Complexes **4b** and **4c** were also shown to possess potent oral activity in this model. All three new complexes have octanol—water partition coefficient (LogP) values in the range of +4, indicating high lipid solubility, while **1** and related catalysts possess highly negative values (Table 1).<sup>30</sup> Both **4b** and **4c** were designed to divert or block the potential metabolic hydroxylation para to the chelated phenol



**Figure 4.** Time-dependent development of carrageenan-induced thermal hyperalgesia in rats (n = 3) was blocked by oral administration of **4a** (100 mg/kg, n = 5) (n = number of animals). Results are expressed as means  $\pm$  standard errors of the mean and were analyzed by two-way ANOVA with Bonferroni post hoc tests; **\*\*** denotes P < 0.05 and **\*\*** denotes P < 0.001 vs vehicle.

through methyl and fluoro substitution, respectively. Full SAR studies incorporating further electron-donating and -withdrawing functionalities will be the subject of future reports.

In conclusion, the results presented herein demonstrate the modular synthesis of a new class of orally active Mn(III) complexes that function as peroxynitrite scavengers. These studies suggest that this new complex design may afford improved in vivo performance through the two-electron reduction of peroxynitrite to nitrite.

## ASSOCIATED CONTENT

**Supporting Information.** Experimental procedures; characterization data; LC-MS, NMR, and UV-vis data; and crystallographic data (CIF). This material is available free of charge via the Internet at http://pubs.acs.org.

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#### REFERENCES

(1) Batinic-Haberle, I.; Reboucas, J. S.; Spasojevic, I. Antioxid. Redox Signaling **2010**, *13*, 877.

(2) Ferrer-Sueta, G.; Radi, R. ACS Chem. Biol. 2009, 4, 161.

(3) Szabo, C.; Ischiropoulos, H.; Radi, R. Nat. Rev. Drug Discovery 2007, 6, 662.

(5) Salvemini, D.; Neumann, W. L. Trends Pharmacol. Sci. 2009, 30, 194.

(6) Salvemini, D. Arch. Biochem. Biophys. 2009, 484, 238.

(7) Groves, J. T.; Marla, S. S. J. Am. Chem. Soc. 1995, 117, 9578.

(8) Stern, M. K.; Jensen, M. P.; Kramer, K. J. Am. Chem. Soc. 1996, 118, 8735.

(9) Lee, J.; Hunt, J. A.; Groves, J. T. *J. Am. Chem. Soc.* 1998, *120*, 6053.
(10) Ferrer-Sueta, G.; Vitturi, D.; Batinic-Haberle, I.; Fridovich, I.; Goldstein, S.; Czapski, G.; Radi, R. *J. Biol. Chem.* 2003, *278*, 27432.

(11) (a) Mahammed, A.; Gross, Z. Angew. Chem., Int. Ed. 2006, 45, 6544. (b) Kupershmidt, L.; Okun, Z.; Amit, T.; Mandel, S.; Saltsman, I.; Mahammed, A.; Bar-Am, O.; Gross, Z.; Youdim, M. B. J. Neurochem. 2010, 113, 363. (c) Gershman, Z.; Goldberg, I.; Gross, Z. Angew. Chem., Int. Ed. 2007, 46, 4320.

(12) (a) Okun, Z.; Kupershmidt, L.; Amit, T.; Mandel., S.; Bar-Am, O.; Youdim, M. B. H.; Gross, Z. ACS Chem. Biol. **2009**, *4*, 910. (b) Kanamori, A.; Catrinescu, M.-M.; Mahammed, A.; Gross, Z.; Levin, L. A. J. Neurochem. **2010**, *114*, 488. (c) Haber, A.; Mahammaed, A.; Fuhrman, B.; Volkova, N.; Coleman, R.; Hayek, T.; Aviram, M.; Gross, Z. Angew. Chem., Int. Ed. **2008**, *47*, 7896.

(13) Drel, V. R.; Pacher, P.; Vareniuk, I.; Pavlov, I.; Ilnytska, O.; Lyzogubov, V. V.; Tibrewala, J.; Groves, J. T.; Obrosova, I. G. *Eur. J. Pharmacol.* **2007**, *569*, 48.

(14) Vareniuk, I.; Pavlov, I. A.; Drel, V. R.; Lyzogubov, V. V.; Ilnytska, O.; Bell, S. R.; Tibrewala, J.; Groves, J. T.; Obrosova, I. G. *Exp. Neurol.* **2007**, 205, 425.

(15) Kim, H.; Burghart, A.; Welch, M. B.; Reibenspies, J.; Burgess, K. Chem. Commun. 1999, 1889.

(16) Burghart, A.; Kim, H.; Welch, M. B.; Thoresen, L. H.; Reibenspies, J.; Burgess, K. J. Org. Chem. **1999**, *64*, 7813.

(17) May, D. A., Jr.; Lash, T. D. J. Org. Chem. 1992, 57, 4820.

(18) (a) Lindsey, J. S.; Schreiman, I. C.; Hsu, H. C.; Kearney, P. C.; Marguerettaz, A. M. J. Org. Chem. **1987**, 52, 827. (b) Littler, B. J.; Ciringh, Y.; Lindsey, J. S. J. Org. Chem. **1999**, 64, 2864.

(19) Parhi, A. K.; Kung, M. P.; Ploessl, K.; Kung, H. F. Tetrahedron Lett. 2008, 49, 3395.

(20) Loudet, A.; Bandichhor, R.; Burgess, K.; Palma, A.; McDonnell, S. O.; Hall, M. J.; O'Shea, D. F. *Org. Lett.* **2008**, *10*, 4771.

(21) Similar behavior was observed for the Mn(V)O corrolazines. See: (a) Mandimutsira, B. S.; Ramdhanie, B.; Todd, R. C.; Wang, H.; Zareba, A. A.; Czernuszewicz, R. S.; Goldberg, D. P. *J. Am. Chem. Soc.* **2002**, *124*, 15170. (b) Lansky, D. E.; Mandimutsira, B. S.; Ramdhanie, B.; Clausen, M.; Penner-Hahn, J.; Zvyagin, S. A.; Telser, J.; Kryzstek, J.; Zhan, R.; Ou, Z.; Kadish, K. M.; Zakharov, L.; Rheingold, A. L.; Goldberg, D. P. *Inorg. Chem.* **2005**, *44*, 4485. (c) Goldberg, D. P. *Acc. Chem. Res.* **2007**, *40*, 626.

(22) Uppu, R. M.; Pryor, W. A. Anal. Biochem. 1996, 236, 242.

(23) Liu, H. Y.; Yam, F.; Xie, Y. T.; Li, X. Y.; Chang, C. K. J. Am. Chem. Soc. 2009, 131, 12890.

(24) Sikora, A.; Zielonka, J.; Lopez, M.; Joseph, J.; Kalyanaraman, B. *Free Radical Biol. Med.* **2009**, *47*, 1401.

(25) (a) Hunt, J. A.; Lee, J.; Groves, J. T. *Chem. Biol.* 1997, 4, 845.
(b) Differences in pH (7.2 vs 7.4) for the boronate assay vs the stopped-flow experiments are in part responsible for the minor differences in the kinetic data.

(26) (a) Lieb, D.; Zahl, A.; Shubina, T. E.; Ivanovic-Burmazovic, I. *J. Am. Chem. Soc.* **2010**, *132*, 7282. (b) Hoogenraad, M.; Ramkisoensing, K.; Driessan, W. L.; Kooijman, H.; Spek, A. L.; Brouwman, E.; Haasnoot, J. G.; Reedijk, J. *Inorg. Chim. Acta* **2001**, *320*, 117.

(27) Lodge, K. J. Chem. Eng. Data 1999, 4, 1321.

(28) For measurement of paw-withdrawl latency from a thermal stimulus (paw withdrawl from a warm surface), see: Hargreaves, K.; Dubner, R.; Brown, F.; Flores, C.; Joris, J. *Pain* **1988**, *32*, *77*.

(29) Salvemini, D.; Wang, Z. Q.; Bourdon, D. M.; Stern, M. K.; Currie, M. G.; Manning, P. T. *Eur. J. Pharmacol.* **1996**, 303, 217.

(30) (a) Kos, I.; Benov, L.; Spasojevic, I.; Reboucas, J. S.; Batinic-Haberle, I. *J. Med. Chem.* **2009**, *52*, 7868. (b) Batinic-Haberle, I.; Ndengele, M. M.; Cuzzocrea, S.; Reboucas, J. S.; Spasojevic, I.; Salvemini, D. *Free Radical Biol. Med.* **2009**, *46*, 212.

(4) Salvemini, D.; Neumann, W. Life Sci. 2010, 86, 604.